AMENDMENTS TO THE SPECIFICATION:

Please replace the paragraph beginning at page 2, line 11, with the following amended paragraph:

The following are examples of immunostimulating oligonucleotides containing a CpG motif, which have been reported as being effective in activating NK cells, wherein the underlined parts show a CpG motif and the parts with capital letters denote a thiolated DNA (Iho, S., and Yamato, S., Annual Review Immunity, 2001, 137-146 (2002): Non-patent Reference 4):

accgataccggtgccggtgacggcaccacg (SEQ ID NO 7)
accgatagcgctgccggtgacggcaccacg (SEQ ID NO 8)
accgatgacgtcgccggtgacggcaccacg (SEQ ID NO 9)
accgattcgcgagccggtgacggcaccacg (SEQ ID NO 10)
ggggggggggggcgatcgggggggggggg (SEQ ID NO 11)
ggggggggggacgatcgtcgggggggggg (SEQ ID NO 12)
ggggggggggggaacgttgggggggggggg (SEQ ID NO 13)
GAGAACGCTCGACCTTCGAT (SEQ ID NO 14)
TCCATGACGTTCCTGATGCT (SEQ ID NO 15)
TCTCCCAGCGTGCGCCAT (SEQ ID NO 16)
GGggtcaacgttgaGGGGGg (SEQ ID NO 17)

Please amend the paragraph beginning at page 7, line 2, as follows:

FIG.2 shows the amino acid sequences of R8 peptide (SEQ ID NO 2) and RGD peptide (SEQ ID NO 3) for use in the present invention.

Please amend the paragraph beginning at page 15, line 10, as follows:

Triple helix schizophyllan was prepared in accordance with the conventional method as described in the literature reference: Schizophyllum commune. Fies (ATCC 44200) available from ATCC (American Type Culture Collection), 10801 University

Boulevard, Manassas, VA, 20110-2209, was subjected to a stationary culture in a minimal medium for seven days. After removal of the cellular materials and insoluble residues, the supernatant was subjected to a supersonic treatment to yield schizophyllan with a triple helix structure having a molecular weight of 450000.

Please amend the paragraph beginning at page 24, line 5, as follows:

As CpG DNA is negatively charged owing to the phosphoric acid groups, it migrates electrophoretically to the anode. Such migration occurs by passing through the network structure of the matrix gel, and therefore the formation of a complex of CpG DNA and schizophyllan reduces the mobility owing to increased molecular weight. Thus, with respect to the complexes of CpG DNA with schizophyllan or chemically modified schizophyllan prepared in the manners as described in Examples 1 and 8, the mobilities were evaluated by electrophoresis, in which the complexes were rendered to migrate on 2% agarose gel in MOPS buffer (20mM MOPS (pH 7.0), 5mM sodium acetate, 1mM EDTA, 3% dimethyl sulfoxide) for one hour at a voltage of 2v/cm. The gel was stained with ~~Gel-Ster~~ GELSTAR™ Nucleic Acids Stain (BMA) and visualized on a transilluminator.

Please amend the two paragraphs beginning at page 24, line 22, as follows:

The Isolation of the mouse-derived intraperitoneal macrophage was carried out in the ordinary manner as described in the literature reference. Thus, a female Balb/c mouse 8 weeks of age was sacrificed by bleeding from the carotid artery. Following sterilization with 70% ethanol, the abdominal skin was cleaved to expose the peritoneum. Cold PBS (phosphate buffered saline) 5ml was injected into the peritoneum, followed by sufficient massage to harvest the fluid. Centrifugation was performed at 1,000 rpm for ten minutes at 4°C using a polypropylene tube. After removal of the supernatant, the resultant was suspended in RPMI1640™ medium containing 10% fetal bovine serum ("New Biochemical Experiments 12: Molecular Immunology I, Immunocytes·Cytokines" edited by Biochemical Society of Japan, published by Tokyo Kagakudojin (1989): Non-patent Reference 22).

The thus obtained macrophage cells 2×10^5, having been suspended in 100μl of the PPMI1640™ medium containing 10% fetal bovine serum, were seeded into a 96-well plate and cultured under 5% CO_2 at 37°C for two hours, so as to render the cells adhered to the plate. To the resultant were added CpG DNA and the complex of CpG DNA and schizophyllan or chemically modified schizophyllan as prepared in Examples 1 and 8, which had been subjected to ultrafiltration (exclusion limit: 3000) to remove the DMSO and readjusted with respect to the concentration. Culturing was conducted at 37°C under 5% CO_2 for 24 hours, followed by the recovery of the culture supernatant.

Please amend the paragraph beginning at page 26, line 6, as follows:

The production of IL-12 cytokine from mouse-derived intraperitoneal macrophage was evaluated in the same manner as in Example 10, by using a sequence of ATG AGC TTC CTG ATG CT, which has phosphorothioate bonds and does not contain any sequence of cytosine-guanine dinucleotide (CpG) (i.e., is not immunostimulating), with the 3′ terminus thereof being linked with forty dA's (hereinafter designated as non-CpG DNA: SEQ ID No. [[2]] 18), in place of CpG DNA as used in Example 10. The results are shown in FIG. 6 (Y. Aramaki, et. al., Biol. Pharm. Bull., 25(3), 351-355 (2002): Non-patent Reference 23).